BRIEF COMMUNICATION

PISONIA GRANDIS DOES NOT APPEAR TO HARBOUR FUNGI KNOWN TO INVADE SEA TURTLE NESTS AT HERON ISLAND, EASTERN AUSTRALIA

Hatching success of loggerhead sea turtle nests is significantly lower at Heron I. (23°26' S, 151°55' E-Capricorn Group, southern Great Barrier Reef), than on the adjacent mainland'. Fungal invasion appears to play a major role in inter-specific and inter-habitat variation in egg mortality between loggerhead (*Caretta caretta* L.) and green (*Chelonia mydas* L.) sea turtles at coral cay and mainland rookeries', and hawksbill (*Eretmochelys imbricata* L.) and flatback (*Natator depressus* Garman) (urtles at other major rookeries in eastern Australia^{*}).

The fungi *Fusarium* oxysporum Schlecht., *E* solani (Mart.) Sace, and *Pseudallescheria boydii* Negroni and Fischer have been isolated from failed turtle eggs at Heron 1.² *Pseudallescheria boydii* is an opportunistic infectant of humans and other animals¹ but there is no record of its being involved in plant disease. However, numerous strains of *F*. oxysporum are wilt pathogens and *F* solani may cause root rol, canker and wilts⁴. Since one of the most distinguishing features of Heron 1, is the dense, central *Pisania grandis* R. Br. forest, it seems possible that this inight be acting as a reservoir for anthracnose fusaria which are also able to invade sea furtle nests.

In its wild state P. grandts (Nyctaginaceae) isalmost exclusively confined to small uninhabited islands with large seabird colonies50, throughout the Indian and Pacific Oceans^{5,6,7,8,9}. In the Capricoust Bunker group of the southern Great Barrier Reef, P. grandis is found on all of the islands. A central forest is usually surrounded by natural fringing vegclation, although erosion may bring the forest to the beachfront". The presence of such forests appears heavily reliant upon abundant seabirds and a specific soil and rock base69. The Jemo Series10 are richly organic, acidic, phosphatic, solls in association with a hardpan or coral conglomerate transformed into calcium phosphate¹¹. This edaphic condition occurs only on coral and coral debris beneath bird colonics8 and is almost exclusive to forests dominated by P. granilis".

Pisonia grandis is often associated with islands hosting pigeons, gannets (Sula spp.) or noddy terns (Auous spp.). If the bird colonies desert the Islands, for whatever reason, the P. grandis forest disappears as it seems unable to survive without the phosphate enriched soil⁶⁷ that aids germination and early devel opment⁶. It is believed P. grandis utilises seabirds for epizoic dispersal⁶, although this has been disputed¹⁷.

Previously, the only fungus associated with Pgrandis at Heron I, was an unidentified basidiomycete ectomycorrhizal symbiont¹⁵. This fungus appears to be unique to P, grandis or at least have a limited host range¹⁴, and could not be one of the three turtle nest mycoflora reported² as none of these is a basidiomycete.

To determine whether *P. grandis* harboured any of the fungal species isolated from failed eggs in sea furtle nests, five individual *P. grandis* trees at the Heron Island Research Station, whose foliage showed anthracnoses, were examined. Two leaves from each tree were collected and washed with sterile, distilled water to remove bird guano before refrigerated storage, Leaf fragments (Lem²) were surface sterilised in 1% AgNO₃ for 2 min then rinsed in 5% NaCl for I min. A final wash in sterile distilled water for 2 min was undertaken to remove any resid ual silver cations. Fragments were cultured as a central moculum on half-strength Potato Dextrose Agar at 28° C for 7 days prior to identification.

Colletotrichum gloeosporioides (Penz.) Penz. and Sacc. was isolated from all leaf fragments with leaf spots. Culture of unblemished fragments did not result in any fungal growth. Colletotrichum is one of the most important genera of plant pathogenic fungi worldwide¹⁵ and can affect stems, shoots, fruit, pods, flowers and leaves¹⁰. It has not been isolated from failed sea turtle eggs and so it seems unlikely that the *P. grandis* forest of Heron Island is hosting fungi likely to have an adverse effect on sea turtle nests.

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^{* 3.} D. Purifort (anpub).

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³ **Rippon, J. W.** (1982). "Medical Mycology. The Pathogenic Fungi and the Pathogenic Actinomycetes 2nd Edn" (W. B. Saunders and Co., Philadelphia).

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⁸ Hunt, D. R. (1967) Kew Bull, 21, 251.

⁹ Cribb, A. B. (1969) Qld Nat. 19, 110-114.

¹⁰ Fosberg, F. R. (1954) Soil Sci. 78, 99-107.

¹¹ Christophersen, E. (1927) Bishop Mus. Bull. 44, 1-79.

¹² Walker, T. A. (1991) Atoll Res. Bull. 350, 1-23.

¹³ Ashford, A. E. & Allaway, W. G. (1982) New Phytol. **90**, 511-519.

¹⁴ Čairney, J. W. G., Rees, B. J., Allaway, W. G. & Ashford, A. E. (1994) *Ibid.* **126**, 91-98.

¹⁵ **Sutton, B. C.** (1992) The Genus *Glomerella* and its anamorph *Colletotrichum* pp. 1-26 *In* Bailey, J. A. & Jeger, M. J. (Eds) "Colletotrichum: Biology, Pathology and Control" (CAB International, Wallingford).

¹⁶ **Dodd, J. C., Estrada, A. & Jeger, M. J.** (1992) Epidemiology of *Colletotrichum gloeosporioides* in the tropics pp. 308-325 *Ibid.*

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